



CHALLENGES AND STRATEGIES IN FRUIT JUICE PROBIOTICIZATION: A REVIEW

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Abstract

Probiotic cultures are successfully applied in various types of food matrices. Numerous food products including dairy, meats, beverages, cereals, vegetables and fruit are utilized as delivery vehicles for probiotics. Fruits are exhibited as appropriate for probiotic products as they don't have any dairy allergens that may prevent usage by part of the population. Also, they contain numerous functional food components like as minerals, vitamins, dietary fibers, and antioxidants. Even though the fruit juices contain various vital nutrients (minerals, vitamins, dietary fibers, antioxidants), there are some strong factors that could limit probiotic survival in juices like food parameters (titratable acidity, molecular oxygen, water activity, presence of salt, sugar, pH and chemicals, like hydrogen peroxide, bacteriocins, artificial flavoring and coloring agents), processing parameters (heat treatment, incubation temperature, cooling rate, packaging materials and storage methods, oxygen levels, volume) and microbiological parameters (strains of probiotics, rate and proportion of inoculation). Different authors proposed successful strategies to improve the survival of probiotics in juices. An easy way to improve probiotic stability in fruit juice could be the fortification of juice with some prebiotics. Induction of Resistance could be achieved through two different strategies: strain cultivation in a lab medium containing different amounts of fruit juices (up to 50%) or added with vanillic acid (phenol stress) or acidified to pH 5.0 (acid stress). These approaches resulted in a prolongation of the viability of lactic cultures by 5 (phenol stress) or 11 days (pH stress).

Introduction

Foods are not only designed to fulfil hunger and to supply necessary nutrients for humans but also, to prevent or reduce the advancement of nutrition-related diseases and to boost physical and mental well-being (Shori, 2014). With respect to this, functional foods play an important role. The term functional food was first utilised in Japan, where the concept of food designed to be medically beneficial to the customer evolved during the 1980s (Siro *et al.*, 2008). Functional food can be defined as food or dietary components that may provide a health benefit beyond the basic function of providing nutrients (Cencic and Chingwaru, 2010). Probiotic foods are one amongst the fastest growing field of functional food production. Probiotics are live microorganisms that when consumed in sufficient amounts exert their health benefits. Probiotic cultures are successfully applied in various types of food matrices. Numerous food products including dairy, meats, beverages, cereals, vegetables and fruit are utilized as delivery vehicles for probiotics. Fruits are exhibited as appropriate for probiotic products as they don't have any dairy allergens that may prevent usage by part of the population. Also, they contain numerous functional food components like as minerals, vitamins, dietary fibers and antioxidants. Juice is

defined as “the extractable fluid contents of tissues or cells.” Each juice has particular, chemical, nutritional and sensorial characteristics, depending upon the kind or nature of fruit or vegetable used. In recent years, studies which were carried out on non-dairy probiotic beverages such as tomato, cabbage, blackcurrant, orange, beet root and carrot juices are performed in combination with different probiotic strains showed the appealing results (Naga *et al.*, 2014). Consumer's interest in whole foods with enhanced nutritional qualities is at an all-time high, and more consumers are opting foods on based on their health benefits (Joao *et al.*, 2012). Fermented foods are food substrates that are invaded or overgrown by edible microorganisms whose enzymes, mainly amylases, proteases and lipases, hydrolase polysaccharides, proteins and lipids to non-toxic products with flavor's, aromas and textures pleasant and attractive to the consumers (Steinkraus, 1997). Number of health benefits is linked to the consumption of probiotic products like as treatment of diarrhea, alleviation of symptoms of lactose intolerance, reduction of blood cholesterol, treatment of irritable bowel syndrome, and inflammatory bowel disease, anti-carcinogenic properties, synthesis of vitamins, and enhancing immunity (Kerry *et al.*, 2018). The market of functional foods is identified by an

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increasing trend; some researchers reported that probiotic foods represent ca. 60-70% of functional foods (Kidist *et al.*, 2019). Generally, the concentrations of 10^6 and 10^7 - 10^8 cfu mL⁻¹ (cfu g⁻¹), respectively, are accepted because of the minimum and satisfactory levels (Karimi *et al.*, 2011).

Challenges for Probiotic Survival in Fruit Juice

Even though the fruit juices contain various vital nutrients (minerals, vitamins, dietary fibers, antioxidants), there are some strong factors that could limit probiotic survival in juices like food parameters (titratable acidity, molecular oxygen, water activity, presence of salt, sugar, pH and chemicals, like hydrogen peroxide, bacteriocins, artificial flavoring and coloring agents), processing parameters (heat treatment, incubation temperature, cooling rate, packaging materials and storage methods, oxygen levels, volume) and microbiological parameters (strains of probiotics, rate and proportion of inoculation) (Perricone *et al.*, 2015).

Mostly, pH exerts a negative effect, but protein and dietary fiber could protect cells from acidic stress; the role of citric and malic acids is divisive, as they appeared to protect probiotics, whereas phenols could cause a much viability loss. Even if the pH is a problem for probiotic survival in juices, Ranadheera *et al.* (2014) assumed that the addition of lactic acid bacteria into fruit juices with low pH may improve the resistance of bacteria to consequent stressful acidic conditions, as those found in GIT.

Strategies for Improving Survival of Probiotic Strains

Different authors proposed successful strategies to improve the survival of probiotics in juices; in this section there is a focus on some case-studies dealing with interesting solutions. An easy way to improve probiotic stability in fruit juice could be the fortification of juice with some prebiotics (dietary fiber, cellulose) or with some ingredients able to exert a protective effect. Adaptation and Induction of Resistance could be achieved through two different strategies: strain cultivation in a lab medium containing different amounts of fruit juices (up to 50%) or added with vanillic acid (phenol stress) or acidified to pH 5.0 (acid stress). These approaches resulted in a prolongation of the viability of lactic cultures by 5 (phenol stress) or 11 days (pH stress). The viability of probiotic bacteria in juices is negatively related to storage temperature, as refrigeration could assure a longer survival, whereas a thermal abuse could show a detrimental effect. Some authors proposed different strategies to fight against the effects of a thermal abuse. Micro encapsulation technologies have been designed and successfully applied using various matrices to protect the bacterial cells from the damage caused by the external environment. Few more inventions in the way of improving viability of probiotic cell count in fruit juices given below.

Sensory Characteristics and Consumer Acceptance of Probiotic Fruit Juice

Many researchers showcased that overall acceptance remained unaffected due to the probiotication of fruit juice like Rodrigues *et al.* (2009) for a fresh apple beverage fermented by *Lb. casei*, Perricone *et al.*, (2014) for pineapple juice containing *Lb. reuteri* and Ellendensen *et al.*, (2012) for apple juice fermented with *Lb. casei*. Luckow and Delahunty (2004) observed that the sensory characteristics of probiotic black currant juice were perfumery and dairy in odour and sour and savoury in flavour. Wunwisa and Kamolnate (2010) found that the addition of probiotic beads enhanced the sensory properties of the product by increasing the swallowing difficulty and remaining particles of the products. The orange and grape juices containing probiotic beads (82.3 and 84.3%, respectively) was accepted by the maximum consumers. Pineapple with Consortium (*Pediococcus pentosaceus* LaG1, *Pediococcus pentosaceus* LBF2 and *Lactobacillus rhamnosus* GG was preferred over the single culture pineapple juice (Tayo and Akpeji, 2016). Luckow *et al.* (2006) explicit that the incorporation of tropical fruit juices, principally pineapple, however conjointly mango or passion fruit (10% v/v), may completely contribute to the aroma and flavor of the end product and may avoid the identification of probiotic off-flavors by consumers. Ranadheera *et al.* (2014) underlined that some fruit juices might naturally mask the “medicinal taste of probiotics.

Safety Issues and Assessment of Probiotic

Probiotics are more often considered as food supplements, and not pharmaceuticals, which suggests the avoidance of extremely thorough testing which are obligatory for all pharmaceuticals. If the probiotic manufacturer makes any specific health claim, probiotics will be categorized as food supplements, meaning that the focus on safety could be underestimated. Furthermore, if any health claims found on the packaging of probiotics, regulatory bodies will examine mainly the validity of such claims and not the safety of the product (De Simone, 2019).

Most probiotic bacteria are marketed in foodstuffs or feed supplements hence probiotic bacteria safety is very important. Through a long research of safe use in food as starter cultures these microbes are confirmed as safe. Bacteria like *Lactobacillus*, *Leuconostoc* and *Pediococcus* species have been used in food processing throughout the history. LAB are classified as generally recognized as safe (GRAS), and there were no reports of any harmful effects from the consumption of these bacteria through many processed products (fermented dairy, fermented vegetables etc.) (Naidu *et al.*, 1999).

In the EU, European Food Safety Authority (EFSA) oversees reviewing health claims of probiotics, which are usually stated on the label while the Food Products Directive

and Regulation (2000/13/EU, 178/2002/EC) controls both probiotics and food supplements. The EFSA also issues the Qualified Presumption of Safety (QPS) for various bacterial strains. The word “presumption” is the only criteria linked with the actual true safety of probiotics, meaning that safety valuation is not the primary concern (Binnendijk and Rijkers, 2013).

Many of the clinical case studies reported that the increased usage of probiotic products of lactobacilli did not cause any increase in incidence or frequency of bacteremia in Finland and few reported the adverse effects of it. Naturally, bifidobacteria are the key bacteria in the intestinal tract of breast-fed infants and are believed to contribute to the good health of infants. As the reports of a harmful effect of these microbes on the host are very sporadic so till now, the safety of the bifidobacteria has not been questioned.

Use of probiotic bacteria in ill persons is restricted to the strains and indications with proven effectiveness. A multidisciplinary approach is needed to judge the toxicological, immunological, gastroenterological, pathological, infectivity, the inherent properties of the microbes, virulence factors comprising metabolic activity, and microbiological effects of probiotic strains. Various methods have been established to evaluate the safety of LAB through the use of in vitro studies, animal studies, and human clinical studies.

Safety considerations of probiotic bacteria comprise of antibiotic resistance profiles, infectivity in immune-compromised animal models, toxin production: probiotic bacteria must be tested for toxin production. The EU Scientific Committee on Animal Nutrition has recommended one of the possible schemes for testing toxin production *i.e.*, hemolytic activity, metabolic activities (D-lactate, bile salt de-conjugation), genetic and pathological side effects, epidemiological surveillance of adverse incidents in consumers (post market).

Potential probiotic health risk can be viewed in two ways (Sanders *et al.*, 2016). The first way encompasses the adverse effects of probiotic per se, while the second way is safety concerns, due to undefined quality standards and manufacturing procedures. However, the only standardization of accurate probiotic safety assessment is a retrospective epidemiologic study, accompanied by thorough pharmacological and toxicological post-marketing vigilance of the product, in order to evaluate further probiotic safety (Kothari *et al.*, 2019).

Labelling Requirements for Probiotic Product

Appropriate labelling and health claims are a pre-requisite for the consumer to make an informed choice. Genus, species and strain, the serving size that delivers the effective dose of probiotic bacteria related to the health claim, minimum viable numbers of each probiotic strain at the end of shelf life, an

precise description of the functional effect, as far as is permissible by law with the required scientific evidence, appropriate storage conditions including the temperature at which the product should be stored, corporate contact details for consumer information, safety in the conditions of recommended use and label information must not mislead the consumers to understand that consumption of the food, ingredient or nutrient of such food, can treat, relieve, cure or prevent a disease should be mentioned on the label of probiotic products.

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Table 1: Examples of Fruit and vegetable Probiotic Product

| Product | Probiotic Strain | References |
|---|--|---|
| Apple | <i>Lactobacillus casei</i> , <i>Lactobacillus plantarum</i> ATCC14917 | de Souza <i>et al.</i> , 2012, Li <i>et al.</i> , 2019 |
| Apricot Juice | <i>Bifidobacterium lactis</i> Bb-12, <i>Bifidobacterium longum</i> Bb-46, <i>Lactobacillus casei</i> 01 and <i>Lactobacillus acidophilus</i> La-5 | Bujn <i>et al.</i> , 2018 |
| Banana puree | <i>Lactobacillus acidophilus</i> | Tsen <i>et al.</i> , 2009 |
| Beet Juice | <i>Lactobacillus plantarum</i> , <i>Lactobacillus casei</i> , <i>Lactobacillus delbrueckii</i> | Yoon <i>et al.</i> , 2005 |
| — | <i>Bifidobacterium animalis</i> subsp. <i>lactis</i> BB-12 and <i>Lactobacillus acidophilus</i> LA-5 | de Oliveira Ribeiro <i>et al.</i> , 2019 |
| Blended orange, carrot, apple and Chinese jujube juice | <i>Lactobacillus plantarum</i> CICC20265, <i>Bifidobacterium breve</i> CICC6184 and <i>Streptococcus thermophilus</i> CICC6220 | Xu <i>et al.</i> , 2019 |
| Cabbage Juice | <i>Lactobacillus plantarum</i> C3, <i>Lactobacillus delbrueckii</i> D7 | Yoon <i>et al.</i> , 2006 |
| Carrot and orange juice | <i>Lactobacillus plantarum</i> | Valero-Cases <i>et al.</i> , 2017 |
| Cashew apple juice | <i>Lactobacillus plantarum</i> , <i>Lactobacillus casei</i> | Kaprasob <i>et al.</i> , 2018, Pereira <i>et al.</i> , 2011 |
| Cantaloupe juice | <i>Lactobacillus casei</i> NRRL B-442 | Fonteles <i>et al.</i> , 2012 |
| Cherry juice | <i>Lactobacillus plantarum</i> , <i>Lactobacillus casei</i> , <i>Lactobacillus paracasei</i> and <i>Lactobacillus rhamnosus</i> | Ricci <i>et al.</i> , 2019 |
| Cornelian cherry juice | <i>Lactobacillus plantarum</i> ATCC 14917 | Mantzourani <i>et al.</i> , 2018 |
| Fruit smoothies | <i>Lactobacillus acidophilus</i> LA-5, <i>Bifidobacterium animalis</i> spp. <i>lactis</i> BB-12 | Rodgers, 2008 |
| Honeydew melon juice | <i>Lactobacillus casei</i> NCIMB 4114 | Saw <i>et al.</i> , 2011 |
| Litchi juice | <i>Lactobacillus casei</i> | Zhen <i>et al.</i> , 2014 |
| Mango and guava juice | <i>Lactobacillus casei</i> , <i>Streptococcus thermophilus</i> , <i>Lactobacillus bulgaricus</i> | Maldonado <i>et al.</i> , 2018 |
| Moringa leaves and beetroot beverage | <i>Lactobacillus plantarum</i> , <i>Enterococcus hirae</i> | Vanajakshi <i>et al.</i> , 2015 |
| Orange, grapefruit, black currant, pineapple, pomegranate, cranberry and lemon juice | <i>Lactobacillus plantarum</i> | Nualkaekul <i>et al.</i> , 2012 |
| Orange, pineapple and cranberry juice | <i>Lactobacillus casei</i> DN 114001, <i>Lactobacillus rhamnosus</i> GG, <i>Lactobacillus paracasei</i> NFBC 43338, <i>Bifidobacterium lactis</i> BB-12 | Sheehan <i>et al.</i> , 2007 |
| Passion fruit juice | <i>Bifidobacterium animalis</i> subsp. <i>lactis</i> BB-12 | Dias <i>et al.</i> , 2018 |
| Pear Juice | <i>Lactobacillus acidophilus</i> | Ankoleka <i>et al.</i> , 2012 |
| Pineapple Juice | <i>Lactobacillus plantarum</i> 299V, <i>Lactobacillus acidophilus</i> La5, <i>Bifidobacterium lactis</i> Bb-12 | Nguyen <i>et al.</i> , 2019 |
| Plum Juice | <i>Lactobacillus kefiranofaciens</i> , <i>Candida kefir</i> , <i>Saccharomyces boulardii</i> | Sheela and Suganya, 2012 |
| Pomegranate juice | <i>Lactobacillus plantarum</i> ATCC 14917, <i>Lactobacillus plantarum</i> , <i>Lactobacillus delbrueckii</i> , <i>Lactobacillus acidophilus</i> , <i>Lactobacillus paracasei</i> | Mantzourani <i>et al.</i> , 2019 Mousavi <i>et al.</i> , 2011 |
| Pumpkin juice | <i>Lactobacillus reuteri</i> | Semjonovs <i>et al.</i> , 2013 |
| Sohiong juice | <i>Lactobacillus plantarum</i> MCC 2974 | Vivek <i>et al.</i> , 2019 |
| Star fruit juice | <i>Lactobacillus helveticus</i> L10, <i>Lactobacillus paracasei</i> L26, and <i>Lactobacillus rhamnosus</i> HN001 | Lu <i>et al.</i> , 2018 |
| Table olives | <i>Lactobacillus</i> GG, <i>Lactobacillus paracasei</i> , <i>Lactobacillus plantarum</i> | Lavermicocca <i>et al.</i> , 2005, Hurtado <i>et al.</i> , 2012 |

Table 2: Examples of Cereal Probiotic Products

| Cereal Probiotic | Product Probiotic Strains | Reference |
|--|---|-----------------------------------|
| Barley malt fermented beverage | <i>Weissellacibaria</i> | Zannini <i>et al.</i> , 2013 |
| Beverage from rice, barley, oats, wheat, soy flour and red grape juice | <i>Lactobacillus plantarum</i> 6E and M6 | Coda <i>et al.</i> , 2012 |
| Breadfruit Flour beverage | <i>Lactobacillus plantarum</i> DPC 206, <i>Lactobacillus acidophilus</i> “de Winkel”, <i>Lactobacillus casei</i> Shirota | Gao <i>et al.</i> , 2019 |
| Cashew juice | <i>Lactobacillus casei</i> NRRL B 442 | Pereira <i>et al.</i> , 2011 |
| Fermented beverage from maize and rice | <i>Lactobacillus plantarum</i> , <i>Torulasporelbrueckii</i> , <i>Lactobacillus acidophilus</i> | Freire <i>et al.</i> , 2017 |
| Fermented oat flour | <i>Streptococcus thermophilus</i> TH-4, <i>Lactobacillus acidophilus</i> LA-5 | Duru <i>et al.</i> , 2019 |
| Fermented oat flour beverage | <i>Lactobacillus plantarum</i> | Gupta and Bajaj, 2017 |
| Legume sprouts | <i>Lactobacillus plantarum</i> 299V | Swieca <i>et al.</i> , 2018 |
| maize-based substrate | <i>Lactobacillus paracasei</i> LBC-81, <i>Saccharomyces cerevisiae</i> CCMA 0731, <i>Saccharomyces cerevisiae</i> CCMA 0732 and <i>Pichiakluyveri</i> CCMA 0615 | Menezes <i>et al.</i> , 2018 |
| Malt beverage | <i>Lactobacillus plantarum</i> NCIMB 8826, <i>Lactobacillus acidophilus</i> NCIMB 8821 | Rathore <i>et al.</i> , 2012 |
| Millet-Based Probiotic Fermented Food | <i>Lactobacillus rhamnosus</i> GR-1 and <i>Streptococcus thermophilus</i> C106 | Di Stefano <i>et al.</i> , 2017 |
| Oat based symbiotic drink | <i>Rhizopus oryzae</i> , <i>L. acidophilus</i> | Gao <i>et al.</i> , 2012 |
| Oat-based probiotic drink | <i>Lactobacillus plantarum</i> B28; <i>Lactobacillus reuteri</i> ATCC 55730 | Angelovet <i>et al.</i> , 2006 |
| Peanut milk | <i>Bifidobacterium pseudocatenumulatum</i> G4 | Kabeir <i>et al.</i> , 2009 |
| Soymilk | <i>Lactobacillus acidophilus</i> | Donkor <i>et al.</i> , 2007 |
| Soymilk with apple juice | <i>Lactobacillus acidophilus</i> | Icier <i>et al.</i> , 2015 |
| Wheat based probiotic Beverage | <i>Lactobacillus acidophilus</i> NCDC-14, <i>Lactobacillus acidophilus</i> NCDC-16 | Sharma <i>et al.</i> , 2014 |
| Wheat/rice cereal infant products | <i>Bifidobacterium animalis</i> subsp. <i>lactis</i> BB-12® | Lebos-Pavunc <i>et al.</i> , 2019 |

Table 3: Strategies for Improving Survival of Probiotic Strains

| Examples of Probiotic | Probiotic Strain | Strategy | Reference |
|--|---|--|----------------------------------|
| Beetroot and carrot Juice | <i>Lb. acidophilus</i> | Fortification with brewer's yeast autolysate | Rakin <i>et al.</i> , 2007 |
| Apple Juice | <i>Lb. rhamnosus</i> | Fortification with Glucan | Saarela <i>et al.</i> , 2006 |
| apple cider, orange, and grape juice | <i>Lb. rhamnosus</i> | Fortification with long and short chain Inulin | Jessica and Sharareh, 2018 |
| pineapple, orange, green apple | <i>Lb. reuteri</i> | Adaptation and Induction of Resistance (Acid and Phenol Stress) | Perricone <i>et al.</i> , 2014 |
| blended juice (orange - grape and passion fruit) | <i>B. breve</i> | Adaptation and Induction of Resistance (Acid Stress) | Saarela <i>et al.</i> , 2011 |
| Freeze Dried Strains | <i>Lb. helveticus</i> | Addition of Tea Extract | Nag and Das, 2013 |
| | <i>Lb. helveticus</i> | Addition of Vit E | Gaudreau <i>et al.</i> , 2013 |
| Model Juice | <i>L. rhamnosus</i> HN001, HOWARU <i>B. Lactis</i> HN001 and <i>L. Paracasei</i> LPC 37 | Addition of grape seed extract, green tea extract and vitamin C | Shah <i>et al.</i> , 2010 |
| Banana puree | <i>Lb. acidophilus</i> | Microencapsulation with Ca-alginate | Tsen <i>et al</i> |
| Sapodilla, grapes, orange and watermelon juices | <i>Lb. plantarum in,</i> | Microencapsulation with Na Alginate | Gaanappriya <i>et al.</i> , 2013 |
| orange and apple juices | <i>Lactobacillus rhamnosus, Bifidobacterium longum, L. salivarius, L. plantarum, L. acidophilus, L. paracasei, B. lactis type Bi -04 and B. lactis type Bi-07</i> | Microencapsulation with Na Alginate | Ding and Shah, 2008. |
| tomato juice | <i>Lb. acidophilus</i> | Microencapsulation with Ca alginate | King <i>et al.</i> , 2007 |
| Orange and Apple Juice | <i>Lb. casei</i> | Microencapsulation with Chitosan | Wunwisa and Kamolnate, 2010 |
| mulberry, maoberry, longan, and melon juices | <i>L. casei</i> 01b, <i>L.acidophilus</i> LA5 and <i>B. lactis</i> Bb-12 | Alginate Encapsulation with cashew flower, pennywort, and yanang | Chaikham, 2015 |